Note: This manuscript has been accepted for publication with Regulatory Toxicology and Pharmacology, which after publication, will be the only definitive repository of its content. Copyright and all rights therein are retained by Elsevier as allowed after appropriate disclaimers.

**Range of the Perfluorooctanoate (PFOA) Safe Dose for Human Health: An International Collaboration**

- Lyle D. Burgoon, Raptor Pharm & Tox, Ltd, USA
- Harvey J. Clewell, Ramboll, USA
- Tony Cox, Cox Associates, USA
- Wolfgang Dekant, University of Würzburg (retired), Germany
- Linda D. Dell, Ramboll, USA
- James A. Deyo, Environmental Protection Authority, New Zealand
- Michael L. Dourson, Toxicology Excellence for Risk Assessment, USA
- Bernard K. Gadagbui, Toxicology Excellence for Risk Assessment, USA
- Philip Goodrum, GSI, USA
- Laura C. Green, Green Toxicology LLC, USA
- K. Vijayavel, Cook Medical, USA
- Travis R. Kline, Geosyntec Consultants, USA
- Tamara House-Knight, GHD, USA
- Michael I. Luster, NIOSH (retired), USA
- Therese Manning, Environmental Risk Sciences Pty Ltd, Australia
- Paul Nathanail, LQM, United Kingdom
- Frank Pagone, RHP Risk Management, USA
- Katie Richardson, Senversa, Australia
- Tiago Severo-Peixe, State University of Londrina, Brazil
- Anurag Sharma, Nitte University, India
- James S. Smith, US DoD, USA
- Nitin Verma, Chitkara University School of Pharmacy, Chitkara University Himachal Pradesh, India
- Jackie Wright, Environmental Risk Sciences Pty Ltd, Australia

**Abstract**
Many government agencies and other expert groups have estimated a dose-rate of perfluorooctanoate (PFOA) that would protect human health. Most of these evaluations are based on many of the same studies (whether of humans, laboratory animals, or both), and all note various uncertainties in our existing knowledge with regard to this chemical and its risks to human health. Nonetheless, the values of these various, estimated, safe-doses (known by various names) vary widely, with some being more than 100,000 fold different than others. This sort of discrepancy invites scrutiny and explanation; otherwise what is the lay public to make of this disparity.

The Steering Committee of the Alliance for Risk Assessment (ARA) called for scientists interested in attempting to understand and potentially narrow these disparities. An advisory committee of nine scientists from four countries was selected from nominations received, and a subsequent invitation to scientists internationally led to the formation of three technical teams (for a total of 24 scientists from 8 countries). The teams reviewed relevant information and independently developed ranges for estimated PFOA safe doses. All three teams determined that the available epidemiologic information could not form a reliable basis for a PFOA safe dose-assessment in the absence of mechanistic data that are relevant for humans at serum concentrations seen in the general population. Based instead on dose-response data from five studies of PFOA-exposed laboratory animals, we estimated that PFOA dose-rates 10 to 70 ng/kg-day are protective of human health.

Introduction

The development of a safe, or subthreshold, dose for perfluorooctanoate (PFOA) has been ongoing for several years. In 2002, a suggested value of 4 ug/kg-day was developed by a team of scientists for the State of West Virginia (DEP, 2002). This assessment was subsequently relied on, in part, by the U.S. Environmental Protection Agency (EPA, 2005) in a draft assessment for EPA’s Office of Toxic Substances. Later, EPA (2009) estimated a safe dose of 0.2 ug/kg-day draft assessment for its Office of Water on more recently available dose-response data.

Outside the U.S., other groups were also estimating safe doses for PFOA, including the European Food Safety Authority (EFSA, 2008) and the United Kingdom (COT, 2009), with both estimating a value of 1.5 ug/kg-day.

EPA (2016) revised its assessment by using a 10-fold lower safe dose (thus estimating 0.02 ug/kg-day), and five years later, revised the value again, this time lowering it quite substantially, to 0.0000015 ug/kg-day (EPA, 2021).

---

1 The term "safe" dose is used throughout this text and is intended to represent a dose just below the population threshold. This population threshold is a point in the dose scale where the first adverse effect, that is the critical effect, is anticipated in a sensitive group of humans. The safe dose concept is used variably by health organization worldwide with slightly different definitions. It is more formally defined here as an estimate (with imprecision spanning perhaps an order of magnitude) of a daily exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of adverse effects during a lifetime.
Other authorities, such as the Drinking Water Inspectorate (2021), Health Canada (2018), the European Food Safety Authority (2018), Food Standards of Australian and New Zealand (FSANZ, 2017) and the Agency for Toxic Substances and Disease Registry (ATSDR, 2018) also have developed or revised their safe doses. These various values have been described previously (e.g., Mikkonen et al., 2020). The World Health Organization (2022) has also recently reviewed this information.

Table 1 lists some of these currently estimated safe doses for PFOA. The wide range in estimated values is striking. These values range between 0.0000015 ug/kg-day and 0.16 ug/kg-day. This disagreement among expert groups was noted by the Steering Committee of the Alliance for Risk Assessment (ARA)² as an issue that might be addressed via collaboration of interested and expert scientists.

It was not the intention of this collaboration to conduct a systematic review and evidence integration or otherwise exhaustively review the literature on PFOA, since many authorities have already adequately done this. Nor was it the intention of this work to critique any individual authority’s approach, although presumably not all approaches can be “correct,” insofar as they disagree by orders of magnitude. Of course, there is still much to learn about the underlying mechanisms of PFOA toxicity before we can arrive at maximally informed estimates of a truly safe dose of PFOA to protect human health. The intent of this work is to estimate a plausible range for such a dose now, anticipating that results of future research will refine and improve on current estimates.

Methods

The Steering Committee of the Alliance for Risk Assessment (ARA) solicited nominations from interested scientists and managers in the early fall of 2022 to form an advisory committee that would shepherd the project entitled “The Perfluorooctanoate (PFOA) Safe Dose”.³ After reviewing nominations, an Advisory Committee was selected from nominations received as shown in Appendix 1.

The Advisory Committee assembled a list of relevant publications on PFOA safe dose and opened a call for interested scientists in the late fall of 2022 to participate in an international collaboration to investigate this issue. After nominations from scientists interested in this collaboration were reviewed by the Advisory Committee, three independent teams of scientists were selected as also shown in Appendix 1, assuring that various scientific experts were represented in each team.

The overall objective of each team was to review relevant information and various agency positions on PFOA in order to determine their safe dose ranges. The teams considered the following criteria in their evaluation: known or suspected mode of action (MOA), overall

---
² See: https://tera.org/Alliance%20for%20Risk/ARA_Steering_Committee.htm
³ See: https://www.tera.org/Alliance%20for%20Risk/Projects/pfoatwo.html
consistency in response among studies, coherence between experimental animal and 
epidemiology data, and robustness of the overall dose response. The science teams were 
directed to review and discuss relevant literature and positions independently of each other and 
in the following manner:

• First, focus on evaluating the evidence regarding potential MOAs for PFOA’s reported 
effects and determining whether the available MOA information would support the 
consideration of the endpoint as a critical effect,

• Then focus on determination of the critical studies for one or more of its critical effect(s),

• Finally, focus on the choice of extrapolation method including the choice of uncertainty 
factors.

The initial focus on PFOA’s MOAs for toxicity was considered to be a critical part of the 
problem formulation step for this project (i.e., to identify the range of a safe dose for PFOA). 
This problem formulation acknowledges that better characterization of hazards (and not merely 
hazard identification) includes consideration of weight of evidence for the MOA and its impact 
on dose-response patterns (NRC, 2009 Science and Decisions, Meek et al. 2014). The sequence 
of work was interspersed with periodic Zoom conference calls in which the teams shared and 
discussed their independently developed results and attempted consensus around the various 
focus topics. Most of our conference calls and team discussions occurred between December 
2022 and March 2023.

Results

The results provided below are summarized by the charges given to the three teams. Teams 
worked independently on each charge and then shared results prior to the periodic international 
Zoom meetings.

The teams reviewed assessments (some of which were draft assessments, indicating ongoing 
development of standards or policy) by national authorities and other authoritative sources, 
specifically, the Agency for Toxic Substances and Disease Registry (ATSDR, 2021), the 
European Food and Safety Authority (EFSA 2018, 2020), the US Environmental Protection 
Agency (USEPA 2021, 2022), the World Health Organization (WHO, 2022), Food Standards 
Australia New Zealand (FSANZ, 2017), Health Canada (2018) and the United Kingdom 
Committee on Toxicity (COT, 2022)) After our deliberations had concluded and before the 
publication of this article, new draft documents had been issued by the USEPA (2023) and 
Health Canada (2023).

Consideration of Mode of Action and Epidemiological Evidence

Overall, each of the teams found it difficult to identify potential MOAs for the various effects of 
PFOA because little mechanistic evidence could be found apart from studies related to the 
disruption of lipid and fatty acid processing in the liver, which has been suggested to be 
responsible for many of the liver effects of PFOA observed in rodents (Andersen et al. 2021a). 
These liver effects of PFOA have been shown to involve activation of multiple, related nuclear
receptors including PPARα, PPARγ, CAR, FXR, LXR, and PXR (Andersen et al. 2021a). However, humans and rodents have been shown to have strikingly different responses, both quantitatively and qualitatively, to lipid-related receptor activation. In both species, there is a core response leading to upregulation of a family of genes controlling fatty acid processing; however, in the rat but not the human, there is a secondary pathway controlled by PPARα that makes the cells more responsive to proliferation. Therefore, the relevance of rodent data for the development of a safe dose range for PFOA is somewhat uncertain. Each of the teams concluded that answers to questions regarding the relevance of animal findings and their associated MOAs to humans were most likely to come from additional in vitro dose-response studies with both rodent and human cells, or in experimental animal models that more closely resemble humans.

There was general agreement that the most likely MOAs for PFOA involved fatty acid mimicry. Fatty acids serve several functions in multiple systems of the body including the ability of the cell to maintain normal fatty acid homeostasis. Membrane fluid dynamics due to the insertion of PFOA into plasma membranes was raised as a possible MOA that could possibly be effective at concentrations below those associated with receptor activation. Such fluidity might be expected due to PFOA’s chemical similarity to plasma lipids and limited volume of distribution from the sole clinical study in humans (suggesting quick sequestration). Insertions of PFOA molecules into the membrane without associated hydrogen bonding might make such membranes less efficient, and if given sufficient dose, might be expected to cause a host of effects. While this was considered a plausible hypothesis there is not yet adequate data supporting it. However, a recent study (Kasten-Jolly and Lawrence 2022) that examined the effects of in vitro exposure of human peripheral blood mononuclear cells to 1, 10, or 100 μM PFOA only observed clear effects on immune cells at the highest concentration (41 ug/mL).

Discussion then segued into the widely different choices of critical effect and their tentative MOA evidence among national authorities. The critical effects identified by national and state authorities included liver effects, developmental effects (decreased body weight, delayed ossification), and impaired T-cell dependent antibody response (TDAR). Until recent years, most critical studies were animal toxicological studies. In 2020, EFSA chose a study of vaccine response to tetanus and diphtheria in one year old infants (Abraham et al. 2020) to derive a toxicity value of 0.0006 μg/kg-day based on a tolerable weekly intake (TWI) of 4.4 ng/kg-bw for four PFAS, including PFOA (EFSA, 2020). Most recently, the USEPA (2023) used epidemiological data for quantitative dose-response assessment when deriving the RfD of 0.00003 μg/kg-day for PFOA as part of recent rulemaking for National Primary Drinking Water Regulations for PFAS. The endpoints and studies described as co-critical effects included decreased antibody response to tetanus and diphtheria vaccine boosters in children (Budtz-Jorgensen and Grandjean, 2018), decreased birth weight in infants (Wikström et al. 2019), and increased total cholesterol in the general population (Dong et al. 2019) (Table 1).

---

4 Critical effect is defined here as the first adverse effect, or its known and immediate precursor, that occurs as dose is increased. It is recognized that multiple effects may be critical (occurring at or around the same dose), and that critical effects in experimental animals may not reflect these same effects found or expected in humans. However, if the critical effect is prevented, then it is assumed that all subsequent adverse effects are prevented.
Budtz-Jorgensen and Grandjean (2018) conducted benchmark dose modeling based on a birth cohort epidemiological study of PFAS and vaccine response in the Faroe Islands. The birth cohort analyzed included 401 children born during 1997-2000 (Grandjean et al. 2012). This study reported a 23% decrease in vaccine antibody titer (VAT) counts for serum anti-diphtheria at age seven per two-fold increase in PFOA and a 28% decrease in VAT counts for serum anti-tetanus at age seven per two-fold increase in PFOA at age five years and after adjusting for age, sex, booster type, and the child’s specific antibody concentration at age five years (Grandjean et al. 2012). The geometric mean concentration of PFOA at age 5 years (2002-2005) was 4.1 ng/ml (interquartile range, 3.3 to 5 ng/ml) indicating low variability in exposure. Both the Food Standards Australia New Zealand (FSANZ 2021) and a science panel convened to evaluate immunotoxicity of PFOA (Garvey et al. 2023) reviewed the Faroe Island data in the context of the broader toxicology and epidemiology literature, and concluded that while VAT may be a biomarker of immunomodulation, it is not suitable to establish immune suppression as a critical endpoint for quantitative risk assessment due to the complexity of accounting for a wider range of potential confounders. Currently, the animal and human evidence for associations between PFAS exposure and incidences of infectious diseases is mixed and inconclusive (Antoniou et al. 2022).

Dong et al. (2019) found an approximate 1.5 mg/dL increase in total cholesterol per ng/mL increase in PFOA in cross-sectional studies of NHANES participants from 2003-2017. Wikström et al. (2019) found birth weight in 1533 infants born during 2007-2010 was decreased by approximately 68 grams per ln-unit of PFOA in maternal blood serum. Maternal blood was sampled early in pregnancy and the association between maternal serum PFOA and decreased birth weight in statistically significant in girls, but not boys. Other agencies reviewed epidemiological studies and found consistent associations consistent associations between PFOA in blood and increases in total cholesterol, decreases in birth weight, and decreases in antibody response to vaccine (ATSDR, 2021; EFSA, 2018, 2020). However, many of the epidemiological studies were cross-sectional designs and there remains the possibility that the associations are confounded by physiological determinants of both biomarkers of exposure and effect or that reverse causation explains the observed associations. For example, EFSA had initially derived a provisional TDI for PFOA and PFOS based on increased cholesterol as the critical effect (EFSA 2018). In the final assessment, EFSA (2020) stated that uncertainty had increased regarding a causal association between PFAS and increased cholesterol because of potential confounding by physiological determinants of PFOA serum concentrations and cholesterol via enterohepatic cycling of bile acids. This hypothesis was one of several discussed in a workshop report of potential mechanisms of increased cholesterol in relation to PFAS that included many recommendations for elucidating mechanisms (Anderson et al. 2021).

Similarly, the association between PFOA (and other PFAS) and decreases in birth weight may be confounded by pregnancy hemodynamics. Both plasma volume expansion and an increased glomerular filtration rate in pregnancy lead to increased elimination of PFOA (Verner et al. 2015). Separately, pregnant women with an impaired glomerular filtration rate are more likely to give birth to babies of lower birth weights while also having increased concentrations of PFOA due to impaired kidney filtration. Meta-analyses of birth weight and PFOA (Steenland et al. 2018) reported small summary decreases in birth weight (average of -10.5 g per ng/ml PFOA, or
approximately 0.35 ounces). In sensitivity analyses to evaluate potential bias associated with
timing of maternal blood sampling, Steenland et al. (2018) reported no effect on birth weight
when maternal blood was sampled early in pregnancy while a larger effect on birth weight was
seen when maternal blood was sampled later in pregnancy.

At a population-level, PFOA blood concentrations have decreased substantially over the past 20
years, from median concentrations of 5.2 ng/ml PFOA (95th percentile, 11.9 ng/ml) in the 1999-
2000 cycle of the National Health and Nutrition Examination Survey (NHANES) to 1.47 ng/ml
PFOA (95th percentile, 3.77 ng/ml) in the 2017-2018 cycle of NHANES (CDC, 2022). This
suggests that there is little variation between individuals in what might be considered
“background” exposure to PFOA and these small differences in concentration partially reflect
differences between individuals in the underlying physiological processes that influence uptake,
distribution, metabolism, and excretion as well as actual differences in environmental exposure.

Other recent research is also relevant: Crawford et al. (2023) reported a summary estimate of an
approximate 12% decrease in anti-diphtheria (95% CI -23% to 0%) and an approximate 12% decrease in anti-tetanus (95% CI -24% to 0%) antibodies per two-fold increase in PFOA in children, a smaller effect than that reported by others (Budtz-Jorgensen and Grandjean, 2018;
Grandjean et al. 2012). Porter et al. (2023) and Bailey et al. (2023) each found that PFOA was
not associated with decreased response to COVID-19 vaccinations when using statistical
methods that allowed for the analysis of repeated measures of serum antibody concentrations and
in populations that had larger variability in PFOA blood concentrations than Abraham et al.
(2020) or Budtz-Jorgensen and Grandjean (2018). Bailey et al. (2023) studied members of two
communities in western Michigan where PFAS had contaminated drinking water (geometric mean 10.3 ng/mL PFOA in one community and 1.62 ng/mL PFOA in the second community).
Porter et al. (2023) studied current and retired workers of one facility that manufactured POSF
dependent PFOA concentration was 1.63 ng/ml (75th percentile, 4.54 ng/ml; 95th percentile 31.70
ng/ml. At a population level, epidemiological studies have reported inconsistent associations
between PFOA blood concentrations and risk of infections, infectious diseases (including
hospitalizations) with some studies reporting positive associations (e.g., Dalsager et al. 2021,
Timmerman et al. 2020), most studies reporting null associations (e.g., Ait-Bamai et al. 2020;
Huang et al. 2020; Manzano-Salgado et al. 2019; Grandjean et al. 2020) and one study reporting
a negative association in boys and a positive association in girls (e.g., Fei et al. 2010; Goudarzi et
al. 2017) while other studies reported mixed evidence (Bulka et al. 2021).

As a result, not all critical effects were thought to be relevant to risk assessment intended to
protect human health, especially in the absence of a postulated mode of action linking early
necessary key events to late key events. While observed associations between PFOA blood
concentrations in populations and diminished levels of serum antibodies following immunization
to one or more specific types of vaccines might prompt additional investigation of
immunosuppressive effects, the current serum concentration/antibody level data were not
deemed suitable for developing a safe dose since the assessments were based upon secondary
immune response (response to diphtheria and tetanus boosters), rather than primary, which
contradicts the WHO immunotoxicology guidelines (derived from Van Loveren et al., 1999), as
a reliable quantitative measure of immune function. Moreover, as several team members noted,
it was unclear whether small decreases in antibody response to vaccines are clinically significant because vast inter- and intra-individual human variability in natural vaccine response exists. This variability precludes any definitive statement in the choice of this endpoint as the critical effect. Recently, a SciPinion panel (Garvey et al., 2023) on immunotoxicity of PFOA suggests that the vaccine threshold of 0.1 IU/ml was not helpful for risk assessment since it is a surrogate of protection and basic immunity is presumed at even lower antibody concentrations (WHO, 2009), most recently 0.01 IU/ml.

Clinical effects in many of the other human observational studies, such as increases in cholesterol and decreases in birth weight, were also of small magnitude or imprecisely estimated. Investigators generally reported that these differences were within normal laboratory reference ranges in relation to PFOA blood concentrations and thus might reflect pharmacokinetic bias or reverse causality due to the fatty acid mimicry based on PFOA’s chemical structure (Andersen et al. 2021b). Although cholesterol changes did not appear to be definitive and were deemed not likely to be the critical effect, studying other inflection points or hormetic responses seemed worthwhile. Reverse causality or confounding by physiological determinants of exposure and effect biomarkers may apply to more than one effect.

An argument can be made that small differences in clinical chemistry biomarkers or clinical effects (decreases in antibody concentrations or birth weight or increases in cholesterol) associated with PFOA blood concentrations can lead to a shift in the population distribution of these clinical parameters, and potentially result in a higher proportion of individuals that experience increased risk of clinical disease. The basis of this argument is an assumption that a causal relationship exists between PFOA and clinical disease in the population. However, increases in frequency and occurrence of infectious disease have only been inconsistently associated with PFOA. For example, some studies have reported an increased risk of hypercholesterolemia (cholesterol level of $\geq 240 \text{ mg/dL}$) with PFOA (Steenland et al. 2009; Winquist and Steenland 2014; Lin et al. 2019) while cardiovascular disease has not been increased with PFOA. In general, studies have not found an increased risk of low birth weight ($<2500 \text{ grams}$) or long-term developmental outcomes associated with decreased birth weight. There is currently insufficient evidence of these adverse effects at the population-level. *In vitro* studies with human cells/tissues over a range of relevant concentrations, similar in design to Kasten-Jolly and Lawrence (2022), are critically needed to elucidate potential MOAs for effects reported in epidemiological studies in order to support any reliable assessment of causality.

A final discussion ensued over whether the dose response information was adequate to develop a safe dose range. This question led to discussion of inflection points or potentially hormetic responses that might yield useful information, such as human observational studies showing an increase in cholesterol at mean or median blood concentrations of 1000 ng/ml or less (Sakr et al. 2007a, 2007b, Steenland et al. 2009, Eriksen et al. 2013, Dong et al. 2019) but the sole human clinical study on PFOA showed decreases at blood concentrations of 175,000–230,000 ng/ml (Convertino et al., 2018).

After presentations, clarifying questions and discussion, the consensus positions summarized in Table 2 and shown below were developed:
1. Several MOAs could be envisioned but not enough evidence exists to establish any one of these MOAs with certainty.

2. Some effects appear to be irrelevant for the determination of a safe dose from current epidemiology data, specifically cholesterol changes and vaccine status.

3. Studying inflection points or perhaps hormesis might help resolve why we have 100,000-fold differences in estimated PFOA safe doses internationally. While differences among such groups can often span a range of 3-fold due to differing times of analyses and methods, this large difference in PFOA is clearly not acceptable for informing confident decision-making, nor can all groups be correct.

**Determination of Studies for PFOA’s Critical Effect(s)**

After reviewing the plethora of relevant information, none of the teams independently considered the epidemiology data, composed primarily of observational studies, to be sufficient to determine a clear effect considering the lack of information regarding the mode of action(s). The results from these studies were considered to be not only potentially confounded, with confounding that was not readily quantified, but also to have serum concentrations from unidentified sources of exposure to PFOA that were not significantly different from background in most studies, making it difficult or impossible to assign a clear exposure-response association, much less causation.

Because of these multiple and significant concerns regarding human observational data, all three teams focused on experimental animals for consideration of the critical effect. However, each team independently reached a different conclusion about the critical effect. One team considered monkey studies as most relevant due to the closeness to humans with PPARα activation for potential liver effects and general physiology, and the difficulty in interpretation of rodent developmental effects. Non-adverse liver effects were seen at all the doses tested in monkeys (3, 10, 20, and 30 mg/kg-day). These effects correlated roughly with non-adverse liver effects seen in the human observational studies and was consistent with the sole human clinical study by Elcombe et al. (2013). Although these liver effects were not considered to be adverse in monkeys, mortality was also observed in monkeys at the higher doses leading to a clear No Observed Adverse Effect Level/Low Observed Adverse Effect Level (NOAEL/LOAEL) boundary. One member from this team reached out to the investigators of the monkey studies to ask for any additional data but none were available.

Another team selected rodent developmental studies rather than liver changes, and specifically Lau et al. (2006) as most relevant due to the consistency in response of several rodent species considering the likely MOA involved fatty acid mimicry. Specifically, PFOA has access to mid-chain fatty acid transport, and biliary and renal excretion and resorption. And while such mimicry might be readily handled by organs such as the liver, it might more readily disturb fatty acid homeostasis in the developing organism, thus supporting the selection of developmental effects as the critical, or perhaps co-critical effect. Moreover, PPARα induced liver effects occurred in rodents at about a 10-fold higher dose than those evoking developmental toxicity.

---

5 The human clinical study of Elcombe et al. (2013) is in the same range and showed no overt effects (50 – 1200 mg/week ÷ 7 days ÷ 70 kg ÷ 0.1 – 2.4 mg/kg-day).
The third team did not judge that the liver effects seen in monkeys, or perhaps other species, were appropriate, since the effects seen were not adverse. Nor did this team consider the developmental effects by Lau et al. (2006) appropriate due to statistical issues associated with the study. Rather, this team was of the general opinion that the overall database was insufficient at this time to make a reliable judgment of critical effect and supported this position with the observation that different health agencies around the world have come to very different decisions. While these differences may not be direct evidence for the overall weakness in the database, the WHO (2022) came to the same conclusion. Specifically, the overall database was considered too uncertain to determine a scientifically based judgment of critical effect. Instead, WHO (2022) made a risk management recommendation.

Finally, all three teams did not rely on several potentially relevant studies of PFOA, and after discussion, agreed that the two-generation study by Macon et al. (2011) was not considered reliable for development of a safe dose range because the statistics in this study appeared to be based on pups and not their mothers. Using pups as the basis of the assessment is not in accordance with US EPA (1991) guidelines. In addition, neither Onischenko et al. (2011) nor Koskela et al. (2016) were used because of too few animals and limited doses used in these studies to generate a confident estimate of the NOAEL/LOAEL interface, and furthermore, it was not certain that the statistics were based on the maternal experimental animals.

After these presentations, clarifying questions and discussion, the following consensus positions were developed as summarized in Table 2 and shown below:

1. **Should human studies be used for the development of the critical effect?**
   No, existing human observational studies cannot be used reliably for this purpose. For example, changes in cholesterol appear to have only a small effect at low doses and an opposite effect at higher doses. These studies may support the choice of critical effect with some of the experimental animal work, however.

2. **Should vaccine responses be used for the development of the critical effect?**
   No, existing human observational vaccine findings are not primary immune responses and of questionable clinical relevance. Based on epidemiological study results, it is premature to assume that a population shift in the distribution of antibody concentrations – if one exists – results in increased risk of susceptibility to diseases. Moreover, higher dose worker exposures do not suggest immune responses.\(^6\)

3. **Should experimental animal studies be used for the development of the critical effect?**
   The overall uncertainty in the database, both epidemiology and experimental animal, is sufficient to give pause to the development of a credible critical effect for PFOA. This conclusion is similar to what WHO (2022) found and for the same or similar reasons.

4. **However, in recognition of the importance of managing PFOA potential health risk, and despite the overall difficulties in the experimental animal studies, a provisional approach was explored as follows:**

\(^6\) Experimental animal work indicates some immune toxicity but only at doses higher than those suggested in human observational studies.
Frank toxicity in both monkeys and rats has been observed in a dose related manner. We might be able to tie these effects into other liver and or developmental endpoints. One member volunteered to conduct a Benchmark Dose (BMD) approach on the relevant monkey and rodent studies and send this to all three teams for consideration (information available upon request).

One team member asked participants to critique and improve upon Green and Crouch (2019) who reviewed the basis of Massachusetts Department of Environmental Protection’s Groundwater and Soil Standards for PFOA and PFOS and suggested an alternate animal test model and target endpoint (i.e., monkey liver toxicity) using a BMD approach.

PFOA is the fluorinated version of the naturally occurring caprylic acid. A big difference between these two chemicals is their half-lives in the human body. Considering whether potential long-term toxicity from caprylic acid matches any of the findings with PFOA may prove useful.

Choice of Extrapolation Method

All teams developed a range in the PFOA safe dose. One team decided to build a range in the safe dose based on several studies of developmental effects in mice. The first study was Lau et al. (2006) with a NOAEL of 23 ug/ml for dose dependent growth deficits in offspring. Other studies considered were Onishchenko et al. (2011), Koskela et al. (2015), Loveless et al. (2006), and Macon et al. (2011). The resulting safe dose range from this collection of studies was 0.011 to 0.27 ug/kg-day.

A second team remained of the opinion that the overall database was insufficient at this time to make a reliable judgment of critical effect. Nevertheless, in order to develop a provisional range, this team focused on two mouse studies, specifically the developmental/reproduction study of Abbott et al. (2007) and the immunotoxicity study of DeWitt et al. (2016), with a range in the NOAELs from 0.3 to 0.94 mg/kg-day. The resulting safe dose range was 3 to 9.4 ug/kg-day from these two values. This team also developed a separate range by adjusting the kinetic comparison between mice and humans based on the work of Zhang et al. (2013) to develop a range of 0.3 to 515 ug/kg-day.

The last team considered liver effect as best meeting the criteria laid out initially and that the results in monkey were most relevant due to comparability of PPARα activation for potential liver effects and general physiology with humans, despite the small numbers of animals and some inconsistency with the reported observations. Butenhoff et al. (2002) showed liver weight increases in monkeys and Green and Crouch (2019) developed a benchmark concentration from these data of 19 ug/ml based on data from this study.

Discussion around these various ranges centered on whether the use of a clearance value from human study Zhang et al. (2013), as describe by Campbell et al. (2022), would be a better choice than clearance values from human observational studies described by Lorber and Egeghy (2011). Also discussed was whether the use of a database uncertainty factor would be reasonable, given the large uncertainty in the overall database. Some concern was also expressed over the use of
Onishchenko et al. (2011) and Koskela et al. (2015) due to the small number of experimental animals and potential use of pup-based statistics. Lastly, the large range in the second team’s calculation appeared to be due to conflating the mouse to human uncertainty factor for toxicokinetic variability with the within human uncertainty factor for toxicokinetic variability. Separating these two seemed reasonable to all participants.

The following consensus positions were developed as summarized in Table 2 and shown below:

1. The various positions of the three science teams appear to overlap, so that developing a provisional range in the PFOA safe dose, based on differing experimental animal studies, seemed reasonable. After discussion by all three teams, there was an agreement to develop a range of the safe dose based on liver effects in monkeys and developmental and immunological effects in mice.

2. The use of human data for this exercise was not entertained, consistent with the earlier consensus of all three science teams that the existing human data were not adequate for identifying safe doses.

3. PFOA has an enormous database, but still has some uncertainty, especially in choosing the critical effect largely due to the relevance to humans of mode(s) of action in animals. A factor of 3-fold for this area of uncertainty should be considered.\(^7\)

4. The use of the average clearance value (either mean, median, mode or geometric versions of these) from the Zhang et al. (2013) human study should be used with any of the experimental animal points of departure if in ug/ml of serum, or by comparison with kinetic information from the relevant species if the points of departure are in units of dose. Moreover, the Zhang et al. (2013) also shows human variability that can be used to develop a data-derived value for within human toxicokinetics. A preliminary analysis by Team 1 gave this a value of ~9-fold.

**Development of a Provisional Safe Dose Range**

A specific provisional range in the PFOA safe dose was subsequently developed based on information from the various consensus calls regarding PFOA’s underlying MOA for various effects, its likely critical effect(s), and the extrapolation of experimental or human data to the presumed sensitive subgroup. The range of the PFOA safe dose is provisionally estimated to be 0.01 to 0.07 ug/kg body weight-day (10-70 ng/kg body weight-day) based on points of departure in Table 3 and uncertainty factors from the studies described in Appendix 2.

**Discussion**

PFAS in general, and PFOA in particular, differ from many other chemicals and mixtures for which safe doses have been estimated. Exposure-response data for the two populations that have

\(^7\) After the meeting several members pointed out that a comprehensive two-generation reproductive toxicity study was conducted in Sprague-Dawley Rats by Butenhoff et al. (2004). EPA used this study to help justify a database UF of 1.
been most highly exposed to PFOA are limited in scope. These two PFOA-exposed groups were
(i) workers who manufactured PFOA, and/or were otherwise occupationally highly exposed and
(ii) a small group of end-stage cancer patients who were administered large doses of PFOA as a
cancer chemotherapeutic drug (Elcombe et al., 2013; Convertino et al., 2018). Notably, though,
observations in both such groups fail to indicate that PFOA presents a significant risk of toxicity.

As noted above, the observational epidemiologic data that associate PFOA body burdens in the
general public with various biological endpoints cannot, in our judgment, serve as reliable basis
for safe dose-assessment. These studies were considered to be not only unquantifiably
confounded but also to have exposures that were not significantly different from background,
which makes the interpretation of any association problematic. We recommend that the
reliability of the results from these epidemiological studies are reconsidered after mechanistic
data become available that supports (or argues against) the hypothesized MOAs; however, in the
absence of mechanistic data relevant to humans at serum concentrations seen in the general
population, the uncertainties of the reliability of the human data that show small differences in
clinical biomarkers are substantial.

At present, the best that can be done, we believe, is to rely on dose-response data from PFOA-
exposed laboratory animals. Mice and rats tend to be good models for humans for most
chemicals; but for PFOA, mice and rats are rather less reliable human-models. Monkeys are
much better models; but, of course, the numbers of monkeys that have been PFOA-exposed are
small; and the endpoints that have been examined remain limited. Future research using non-
human primates might well yield useful information for purposes of human health risk
assessment.

With regard to the potential carcinogenicity of PFOA, there was general agreement that the
EPA’s proposed change in the categorization of PFOA from “suggestive evidence” to “likely
carcinogen” is not justified. The EPA’s determination was based primarily on clear evidence of
PFOA-induced liver tumors in rodents and variously published associations between PFOA
concentrations and kidney cancer in humans (Barry et al. 2013, Vieira et al. 2013, Steenland and
Woskie 2012, Shearer et al. 2020), and the EPA identified a case-control study of renal cell
carcinoma (RCC) nested within the screening arm of Prostate, Lung, Colorectal and Ovarian
(PLCO) cancer screening trial study as particularly influential (Shearer et al. 2020).

However, as is well known, rodent liver tumors are observed only at doses associated with
peroxisomal proliferation, a response of limited relevance to human exposures. And, on our
opinion, the relevant epidemiological studies have not adequately considered the potential for
confounding by impaired renal function, which is associated with both PFOA clearance and
kidney cancer.

With regard to kidney cancer, we note that if PFOA were a genuine cause of this cancer-type in
humans, then one might expect that the massive doses of PFOA used in the rodent (and monkey)
bioassays would have also induced kidney tumors. Yet, they did not.
Kidney cancer is frequently associated with impaired renal function and alterations in renal function that resulted in decreased PFOA excretion would result in a consequent increased PFOA concentration in serum. Cross-sectional analyses of adults exposed at background levels (Shankar et al. 2011) and of children exposed at high levels (Watkins et al. 2013) found a positive association between lower kidney function and higher measured serum PFOA. Dhingra et al. (2016), performed an analysis of cross-sectional studies reporting associations between PFOA and renal function, and concluded that pharmacokinetic confounding led to the observed associations. While Shearer et al. (2021) adjusted their results for estimated glomerular filtration rate (eGFR), adjusting for eGFR alone would not adequately control for this potential confounding due to the extensive role of renal transporters in the clearance of PFOA.

The international process described in this brief communication has several advantages. Many of the scientists who volunteered for this task are well published in the area of PFOA, or in one or more of PFOA’s designated critical effects, or in one or more of the extrapolation methods used to determine the provisional range of its safe dose. Many of these scientists are also intimately familiar with one or more of the agency positions on PFOA. Despite these credentials and familiarity, or perhaps because of them, uniformity of thought was not present, at least initially, and the Zoom call meetings were often lively but respectful. Therefore, the eventual consensus of 27 scientists from 8 countries over 6 months can perhaps be afforded a higher degree of trust than position developed with fewer or less diverse viewpoints.

This process, however, also has its drawbacks. First, it depended on group or self-nominations and from individuals from groups that may or may not appreciate a particular agency position. This concern was addressed in two ways. First, nominations to the Advisory Committee were solicited by the Steering Committee from known experts in the field along with an open nomination process. Members were then selected by the Alliance for Risk Assessment Steering Committee after a review of credentials. This Steering Committee is composed of 5 scientists, 3 from governments, one from a university and one from an environmental science non-government organization. In turn, members of the 3 science teams were selected by the Advisory Committee after an open nomination process and review of proffered biographical sketches/resumes. Balances were maintained among affiliations within each science team. A second drawback is that no funding was received for this work, making it difficult to follow-up on nuances of data that needed additional consideration.

The suggested provisional safe dose range of this international collaboration is 0.01 to 0.07 ug/kg-day. This range encompasses the single value of Health Canada (2018) and the projected range of values for the WHO (2022) and lies slightly below the value of Food Standards of Australia and New Zealand (FSANZ, 2017; Australian Government, 2022). However, this range is well above the single values of both EFSA (2020) and EPA (2023). The principal reasons for the larger disparity between this provisional range with these latter two single values is the unanimous judgment of the international collaboration that the existing human data are not sufficiently credible as a basis of the PFOA safe dose in the absence of mechanistic data that are relevant to humans at serum concentrations seen in the general population. In this regard, Health Canada, the WHO and Food Standards of Australia and New Zealand are in agreement with the
Collaboration—the use of human data is not sufficiently credible as the basis for the PFOA safe
dose.

Additional thoughts from other colleagues are welcome. We continue to believe that,

...It is the mark of an instructed mind to rest satisfied with the degree of precision which
the nature of the subject permits and not to seek an exactness where only an
approximation of the truth is possible. Aristotle

Acknowledgements

The development of research and subsequent publication was under the auspices of the Alliance
for Risk Assessment (ARA). No funding was accepted from any organization for this work. The
authors appreciate the insights and leadership of Professor Ravi Naidu of the Contamination
Assessment and Remediation of the Environment, University of Newcastle, Australia.

Disclaimers

The views expressed in this article are those of the authors and do not necessarily reflect the
official policy or position of the Department of the Navy, Department of Defense, nor the U.S.
Government. JS is an employee of the U.S. Government. This work was prepared as part of his
official duties. Title 17, U.S.C., §105 provides that copyright protection under this title is not
available for any work of the U.S. Government. Title 17, U.S.C., §101 defines a U.S.
Government work as a work prepared by a military Service member or employee of the U.S.
Government as part of that person’s official duties.

HC and LD are salaried employees of Ramboll US Consulting, Inc., a consulting firm that
provides scientific and technical support to a variety of clients in private and public sectors.
Participation in the project and the preparation of the manuscript reflects the professional work
of the authors and may not necessarily reflect the views of Ramboll US Consulting, Inc. or its
parent company, Ramboll Group A/S. HC and LD did not receive outside funding to participate
in this project or prepare the manuscript; the manuscript was prepared on their own time or
supported by Ramboll as part of their usual employment responsibilities. Prior to preparing the
manuscript, LD has been retained as an expert witness on behalf of defendants in litigation
matters pertaining to certain PFAS.

MD and BG are employees of Toxicology Excellence for Risk Assessment (TERA), which has
worked over a number of years for governmental and nongovernmental sponsors on PFAS
issues. However, no outside funding was accepted to prepare this manuscript nor to do the
analyses underlying it.

THK is an employee of GHD, Inc., a consulting firm, serving a variety of clients in the private
and public sector. The time spent on this manuscript was performed on the author’s own time.
and was not supported financially by any entity. The views expressed in this article are those of
the author and do not necessarily reflect the position or policy of GHD.

FP is an employee of RHP Risk Management, a consulting firm, serving a variety of clients in
the private and public sector. RHP has performed consulting and testifying work on various
matters including PFAS. Neither FP nor RHP has shared this work with any RHP client nor
elicited input into the design, preparation, or review of this work prior to publication. The time
spent on this manuscript was either supported by RHP or was performed on the author’s own
time.

The views expressed in this article are those of the authors and do not necessarily reflect the
official policy or position of the New Zealand Environmental Protection Authority.

References

Abbott, BD; Wolf, CJ; Schmid, JE; Das, KP; Zehr, RD; Helfant, L; Nakayama, S; Lindstrom,
AB; Strynar, MJ; Lau, C. (2007). Perfluorooctanoic acid induced developmental toxicity in the
mouse is dependent on expression of peroxisome proliferator activated receptor-alpha. Toxicol

Abraham, K.; Mielke, H.; Fromme, H.; Völkel, W.; Menzel, J.; Peiser, M.; Zepp, F.; Willich,
S.N.; Weikert, C. Internal Exposure to Perfluoroalkyl Substances (PFASs) and Biological
Markers in 101 Healthy 1-Year-Old Children: Associations between Levels of Perfluorooctanoic
Acid (PFOA) and Vaccine Response. Arch Toxicol 2020, 94, 2131–2147, doi:10.1007/s00204-
020-02715-4

Agency for Toxic Substances and Disease Registry (ATSDR), 2021. Toxicological Profile
and Human Services.

prenatal exposure to per- and polyfluoroalkyl substances on childhood allergies and common
infectious diseases in children up to age 7 years: The Hokkaido study on environment and
children’s health. Environ Int. 143: 105979.

Alliance for Risk Assessment (ARA, 2022). At:
https://tera.org/Alliance%20for%20Risk/Projects/pfoatwo.html. Last accessed on 4-18-23.

Andersen ME, Hagenbuch B, Apte U, Corton JC, Fletcher T, Lau C, Roth WL, Staels B, Vega GL,
Clewell HJ, Longnecker MP. 2021a. Why is elevation of serum cholesterol associated with exposure to
perfluoroalkyl substances (PFAS) in humans? A workshop report on potential mechanisms. Toxicology,


Butenhoff, J; Costa, G; Elcombe, C; Farrar, D; Hansen, K; Iwai, H; Jung, R; Kennedy, G; Lieder, P; Olsen, G; Thomford, P. (2002). Toxicity of ammonium perfluorooctanoate in male cynomolgus monkeys after oral dosing for 6 months. Toxicol Sci 69: 244-257.


Crawford L, Halperin SA, Dzierlenga MW, Skidmore B, Linakis MW, Nakagawa S, Longnecker MP. 2023. Systematic review and meta-analysis of epidemiologic data on vaccine response in...
relation to exposure to five principal perfluoroalkyl substances. Environ Int. 172:107734. doi:

Dalsager L, Christensen N, Halekoh U, Timmermann CAG, Nielsen F, Kyhl HB, Husby S,
Grandjean P, Jensen TK, Andersen HR. 2021. Exposure to perfluoroalkyl substances during fetal
life and hospitalization for infectious disease in childhood: A study among 1,503 children from
the Odense Child Cohort. Environ Int 149: 106395.

Dewitt, JC; Williams, WC; Creech, NJ; Luebke, RW. (2016). Suppression of antigen-specific
antibody responses in mice exposed to perfluorooctanoic acid: Role of PPARα and T- and B-cell

to determine the associations between per-and polyfluoroalkyl substances and cholesterol: Trend

Regulations 20161 specific to PFOS (perfluorooctane sulphonate) and PFOA
(perfluorooctanoic acid) concentrations in drinking water. London, UK.

Com/24/ee/73/f58267c7d70dde/WO2011101643A1.pdf.

EFSA (European Food and Safety Authority), 2008. Perfluorooctane sulphonate (PFOS),
perfluorooctanoic acid (PFOA) and their salts. Scientific Opinion of the Panel on
Contaminants in the Food chain. EFSA J 653, 1–131.

EFSA (European Food and Safety Authority), 2018. Risk to human health related to the presence
of perfluorooctane sulfonic acid and perfluorooctanoic acid in food. EFSA Panel on
Contaminants in the Food Cain (CONTAM). EFSA J 16 (12), 5194.

EFSA Panel on Contaminants in the Food Chain (EFSA CONTAM Panel), 2020. Risk to human
health related to the presence of perfluoroalkyl substances in food. EFSA J 18(9):6223.

PFOA and PFOS levels and total cholesterol in a middle-aged Danish population. PLoS ONE
8:e56969.

Fei C, McLaughlin JK, Lipworth L, et al. 2010. Prenatal exposure to PFOA and PFOS and risk
10.1016/j.envres.2010.08.004.

18


dose/concentration-response assessment. Geneva Switzerland. Available at

Kasten-Jolly J, Lawrence DA. Perfluorooctanesulfonate (PFOS) and perfluorooctanoic acid
(PFOA) modify in vitro mitogen- and antigen-induced human peripheral blood mononuclear cell
(PBMC) responses. J Toxicol Environ Health A. 2022 Sep 2;85(17):715-737. doi:

Koskela, A; Koponen, J; Lehenkari, P; Viluksela, M; Korkalainen, M; Tuukkanen, J. (2017).
Perfluoroalkyl substances in human bone: concentrations in bones and effects on bone cell

Lau, C; Thibodeaux, JR; Hanson, RG; Narotsky, MG; Rogers, JM; Lindstrom, AB; Strynar, MJ.
90: 510-518.

Lin P, Cardenas A, Hauser R, Gold DR, Kleinman K, Hivet MF, Fleisch AF, Calafat AM,
Webster TF, Horton ES, Oken E. 2019. Per- and polyfluoroalkyl substances and blood lipid
levels in pre-diabetic adults-longitudinal analysis of the diabetes prevention program outcomes
study. Environ Int 129: 343-353.

Lorber, M; Egeghy, PP. (2011). Simple intake and pharmacokinetic modeling to characterize
exposure of Americans to perfluorooctanoic acid, PFOA. Environ Sci Technol 45: 8006-8014.

Comparative responses of rats and mice exposed to linear/branched, linear, or branched
ammonium perfluorooctanoate (APFO). Toxicology 220 (2–3), 203–217.
https://doi.org/10.1016/j.tox.2006.01.003.

Macon, MB; Villanueva, LR; Tatum-Gibbs, K; Zehr, RD; Strynar, MJ; Stanko, JP; White, SS;
Helfant, L; Fenton, SE. (2011). Prenatal perfluorooctanoic acid exposure in CD-1 mice: low-
dose developmental effects and internal dosimetry. Toxicol Sci 122: 134-145.

Manzano-Salgado CB, Granum B, Lopez-Espinosa MJ, Ballester F, Iñiguez, C, Gascón M, Martínez D,
exposure to perfluoroalkyl substances, immune-related outcomes, and lung function in children from a

relevance (species concordance) framework: Evolution of the Bradford Hill considerations and
10.1002/jat.2984.

McMullen PD, Battacharya S, Woods CG, Pendse SN, McBride MT, Soldatow VY, Deisenroth
C, LeCluyse EL, Clewell RA, Andersen ME. Identifying qualitative differences in PPARα
signaling networks in human and rat hepatocytes and their significance for next generation
chemical risk assessment methods. Toxicol In Vitro. 2020 Apr;64:104463. doi:

for improving the characterisation of risk from exposures to per and polyfluorinated alkyl


Onishchenko, N; Fischer, C; Wan Ibrahim, WN; Negri, S; Spulber, S; Cottica, D; Ceccatelli, S.
(2011). Prenatal exposure to PFOS or PFOA alters motor function in mice in a sex-related

Porter AK, Kleinschmidt SE, Andres KL, Reusch CN, Krisko RM, Taiwo OA, Olsen GW,
Longnecker MP. 2022. Antibody response to COVID-19 vaccines among workers with a wide
range of exposure to per- and polyfluoroalkyl substances. Environ Int. 169:107537. doi:
10.1016/j.envint.2022.107537

Post, Gloria. (2021). Recent US State and Federal Drinking Water Guidelines for Per- and
https://doi.org/10.1002/etc.4863.

enzymes in workers with occupational exposure to ammonium perfluorooctanoate. J Occup

enzymes related to a serum biomarker of exposure (ammonium perfluorooctanoate or APFO) as
part of a general health survey in a cohort of occupationally exposed workers. J Occup Environ

SciPinion, 2023. Risk Assessment of PFAS: An international panel of experts provides guidance
on key questions in the risk assessment of PFAShttps://scipinion.com/panel-findings/risk-
assessment-of-pfas/. Last accessed on April 27.

Ammonium Perfluorooctanoate (C8) Assessment Of Toxicity Team (CATT) Report.

Steenland K, Tinker S, Frisbee S, Ducatman A, Vaccarino V. 2009. Association of
perfluorooctanoic acid and perfluorooctane sulfonate with serum lipids among adults living near


Table 1. Safe Doses of PFOA and PFOS from International Authorities

<table>
<thead>
<tr>
<th>Authority</th>
<th>Safe Dose ug/kg-day</th>
<th>Point of Departure (POD_{HED})</th>
<th>Uncertainty Factors</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alliance for Risk Assessment (this paper)</td>
<td>0.01-0.07</td>
<td>Various (see text): 4.35 to 23 ug/ml of serum</td>
<td>Animal-human kinetic factor = 1&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Animal-human dynamic factor = 3&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Human toxicodynamic factor = 3&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Human toxicokinetic factor = 8.4&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Database uncertainty factor = 1&lt;sup&gt;e&lt;/sup&gt;</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Human clearance = 0.23 ml/day-kg&lt;sup&gt;f&lt;/sup&gt;</td>
</tr>
<tr>
<td>European Food Safety Authority (EFSA, 2020)</td>
<td>0.00063&lt;sup&gt;g&lt;/sup&gt;</td>
<td>17.5 ng/mL (BMDL&lt;sub&gt;10&lt;/sub&gt;)</td>
<td>Decreased anti-tetanus and anti-diphtheria antibody concentration</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>None applied</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>BMD derived in sensitive population (infants) and response is risk factor for disease rather than a disease.</td>
</tr>
<tr>
<td>Food Standards Australia/New Zealand (2017)</td>
<td>0.16</td>
<td>4.9 ug/kg-day</td>
<td>Within human variability = 10</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Animal to human extrapolation = 3</td>
</tr>
<tr>
<td>Health Canada (2018)</td>
<td>0.02</td>
<td>0.52 ug/kg-day</td>
<td>Within human variability = 10</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Animal to human extrapolation = 2.5</td>
</tr>
<tr>
<td>US Environmental Protection Agency (2021 DRAFT)</td>
<td>0.0000015</td>
<td>0.0000149 ug/kg-day decreased anti-tetanus antibody concentration</td>
<td>Within human variability = 10</td>
</tr>
<tr>
<td>US Environmental Protection Agency (2023 DRAFT)</td>
<td>0.00003&lt;sup&gt;g&lt;/sup&gt;</td>
<td>Various (human): 0.000305 ug/kg-day (decreased anti-tetanus and anti-diphtheria antibody concentration), 0.000275 ug/kg-day (increased serum cholesterol) 0.000292 μg/kg-day (decreased birth weight)</td>
<td>Within human variability = 10</td>
</tr>
<tr>
<td>Authority</td>
<td>Safe Dose (ug/kg-day)</td>
<td>Point of Departure (POD&lt;sub&gt;HEI&lt;/sub&gt;)</td>
<td>Uncertainty Factors</td>
</tr>
<tr>
<td>------------------------------------------------</td>
<td>-----------------------</td>
<td>----------------------------------------</td>
<td>-----------------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
</tbody>
</table>
| World Health Organization (2022)              | 0.02                  | Estimated based on PFOA water level of 100 ng/liter | • WHO made a risk management call of 100 ng/liter  
• This value can be used to estimate the comparable safe dose of 0.02 ug/kg-day using 2 liters of water consumption per day, a 60 kg body weight and a 20% relative source contribution. |

1 Factor is not needed since PODs are based on serum concentrations.  
2 The use of a 3 is the US EPA default position (USEPA, 2014); the IPCS (2005) default is 2.5.  
3 The use of a 3 is both the US EPA and IPCS default positions.  
4 This value of 8.4 is derived by dividing the value of 0.79 ml/day/kg, which is the arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) by a value of 0.094 ml/day/kg, which is the arithmetic 95% lower bound clearance of sensitive group from Zhang et al. (2013, Table 2).  
5 Data base factor of 1 was considered appropriate for most PODs.  
6 This value of 0.23 ml/day/kg is the geometric mean clearance from Zhang et al. (2013, Table 2) assuming steady state.  
7 Sum of four PFAS: PFOA, PFNA, PFHxS, and PFOS  
8 USEPA, 2023 is DRAFT RfD in response to SAB comments that EPA consider multiple studies of different endpoints in different populations to derive an RfD.
### Table 2. International Collaboration Consensus Statements

<table>
<thead>
<tr>
<th>Consensus on Mode of Action</th>
<th>Several MOAs could be envisioned but not enough evidence exists to establish any one of these MOAs with certainty.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Certain effects appear to be irrelevant for the determination of a safe dose in the absence of mode of action information relevant to humans, specifically differences in cholesterol &amp; vaccine response.</td>
</tr>
<tr>
<td></td>
<td>Studying inflection points or perhaps hormesis might help resolve why we have 100,000-fold differences in the PFOA safe dose internationally.</td>
</tr>
<tr>
<td>Consensus on Critical Effect</td>
<td>Existing human observational studies cannot be used reliably for developing the critical effect in the absence of mechanistic data relevant to humans at serum concentrations seen in the general public.</td>
</tr>
<tr>
<td></td>
<td>Existing human observational vaccine findings are not primary immune responses and not of clinical relevance. Epidemiological studies of risk of infectious diseases have been mixed. In populations with higher PFOA blood concentrations, there response to SARS-CoV-2 antibodies from MRNA vaccines</td>
</tr>
<tr>
<td></td>
<td>The overall uncertainty in the database is sufficient to give pause to the development of a credible critical effect for PFOA. However, in recognition of the importance of managing PFOA potential health risks, a provisional approach could be developed based on several experimental animal studies.</td>
</tr>
<tr>
<td>Consensus on Extrapolation Method</td>
<td>The various positions of the three science teams overlap, so developing a provisional range in the PFOA safe dose, based on differing experimental animal studies, seemed reasonable.</td>
</tr>
<tr>
<td></td>
<td>Human data are not an acceptable basis of the safe dose.</td>
</tr>
<tr>
<td></td>
<td>PFOA has an enormous database, but still has some uncertainty, suggesting that a 3-fold factor may be reasonable.</td>
</tr>
</tbody>
</table>
A clearance value from the Zhang et al. (2013) should be used with any of the experimental animal points of departure and can be used for a data-derived value for human toxicokinetics.
Table 3. Experimental Animal Studies as the Basis of the Provisional Safe PFOA Dose.*

<table>
<thead>
<tr>
<th>Reference</th>
<th>Safe Dose ug/kg-day</th>
<th>Point of Departure (POD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Butenhoff et al. (2002).</td>
<td>0.06</td>
<td><strong>Monkey</strong>: Point of Departure = 19 ug/ml from Green and Crouch (2019) based on a serum PFOA benchmark concentration (BMC) for <em>increased liver weight</em></td>
</tr>
<tr>
<td>Lau et al. 2006</td>
<td>0.07</td>
<td><strong>Mouse</strong>: Point of Departure = 1 mg/kg-day or 23 µg/ml No Observed Adverse Effect Level (NOAEL) for <em>dose-dependent growth deficits</em> for gestation days 1-17</td>
</tr>
<tr>
<td>Loveless et al. (2006)</td>
<td>0.01</td>
<td><strong>Mouse</strong>: Point of Departure = 4.35 µg/ml based on a serum PFOA benchmark concentration by New Jersey/New Hampshire (Post et al., 2021) for <em>lipid parameters/relative liver weight</em> in male mice</td>
</tr>
<tr>
<td>Abbott et al. (2007)</td>
<td>0.03</td>
<td><strong>Mouse</strong>: Point of Departure = 0.3 mg/kg-day (10.4 ug/ml) NOAEL for <em>neonatal survival</em> found</td>
</tr>
<tr>
<td>DeWitt et al. (2016)</td>
<td>0.07</td>
<td><strong>Mouse</strong>: Point of Departure = 0.94 mg/kg-day (no serum values available) NOAEL for <em>immune suppression</em> found</td>
</tr>
</tbody>
</table>

* See Appendix 2 for details of the various calculations.
Appendix 1

Advisory Committee

- Lyle D. Burgoon, Raptor Pharm & Tox, Ltd, USA
- Harvey J. Clewell, Ramboll, Global
- Tony Cox, Cox Associates, USA
- Michael L. Dourson, TERA, USA
- Tamara House-Knight, GHD, Global
- Ravi Naidu, CRC CARE, Australia
- Paul Nathanail, LQM, United Kingdom
- James S. Smith, US DoD, USA
- Nitin Verma, Chitkara University School of Pharmacy, Chitkara University Himachal Pradesh, India

Independent Science Teams

Team 1

- Lyle D. Burgoon, RaptorPharmTox, USA
- Paul Nathanail, LQM, United Kingdom
- Shanon E. Ethridge, International Association for Plumbing and Mechanical Officials Research and Testing, USA
- K. Vijayavel, Cook Medical, USA
- Michael I. Luster, NIOSH (retired), USA
- Therese Manning, Environmental Risk Sciences Pty Ltd, Australia
- Tiago Severo-Peixe, State University of Londrina, Brazil
- Andrea Wojtyniak, Geosyntec, Canada

Team 2

- Harvey J. Clewell, Ramboll, USA
- Tamara House-Knight, GHD, Global
- Linda Dell, Ramboll, Global (MS, Epidemiology; 30 years of experience)
- James A. Deyo, Environmental Protection Authority, New Zealand
- Bernard K. Gadagbui, Toxicology Excellence for Risk Assessment, USA
- Travis R. Kline, Geosyntec Consultants, USA
- Katie Richardson, Senversa, Australia
- Anurag Sharma, Nitte University, India
Team 3

- James S. Smith, NMCPHC, USA
- Nitin Verma, Chitkara University School of Pharmacy, Chitkara University Himachal Pradesh, India
- Wolfgang Dekant, University of Würzburg (retired), Germany
- Philip Goodrum, GSI, USA
- Laura C. Green, Green Toxicology LLC, USA
- Frank Pagone, RHP Risk Management, USA
- Jackie Wright, Environmental Risk Sciences Pty Ltd, Australia
Appendix 2

Monkey: Point of Departure = 19 ug/ml from Green and Crouch (2019) based on a serum PFOA benchmark concentration (BMC) for increased liver weight in Butenhoff et al. (2002).

- Monkey to human toxicokinetic factor = 1 [Factor is not needed since BMD is based on serum concentration]
- Monkey to human toxicodynamic factor = 2.5 [IPCS (2005) default or 3 EPA (2014) default]
- Human toxicodynamic factor = 3 [default of IPCS (2005) and EPA (2014)]
- Human toxicokinetic factor = 8.4 [0.79 ml/day/kg arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) ÷ 0.094 ml/day/kg arithmetic 95% lower bound clearance of sensitive group from Zhang et al. (2013, Table 2)]
- Database uncertainty factor = 1 (Although it could be argued that the small number of animals in the study justifies an additional uncertainty factor; the counter-argument is that these are primates. See also footnote 7.)

- RfD serum concentration = 0.25 ug/ml [19 ug/ml ÷ (1 x 3 x 3 x 8.4 x 1) = 0.25]
- RfD = 0.06 ug/kg-day [0.25 ug/ml x 0.23 ml/day/kg [geometric mean clearance from Zhang et al. (2013, Table 2) assuming steady state]

Mouse: Point of Departure = 1 mg/kg-day or 23 µg/ml No Observed Adverse Effect Level (NOAEL) for dose-dependent growth deficits in the Lau et al. 2006 for gestation days 1-17

- Mouse to human toxicokinetic factor = 1 (Factor is not needed since BMD is based on serum concentration)
- Mouse to human toxicodynamic factor = 2.5 [IPCS (2005) default or 3 EPA (2014) default]
- Human toxicodynamic factor = 3 [default of IPCS (2005) and EPA (2014)]
- Human toxicokinetic factor = 8.4 [0.79 ml/day/kg arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) ÷ 0.094 ml/day/kg arithmetic 95% lower bound clearance of sensitive group from Zhang et al. (2013, Table 2)]
- Database uncertainty factor = 1 (Although it has been argued that problems with this study might justify an additional uncertainty factor; the counter-argument is that US EPA uses a value of 1. See also footnote 7.)

- RfD serum concentration = 0.30 ug/ml [23 ug/ml ÷ (1 x 3 x 3 x 8.4 x 1) = 0.30]
- RfD = 0.07 ug/kg-day [0.30 ug/ml x 0.23 ml/day/kg [geometric mean clearance from Zhang et al. (2013, Table 2) assuming steady state]

Notes:
- It could be argued that the fetal toxicity is secondary to disruption of lipid metabolism in the dam, as evidenced by the increased maternal liver weight at all doses.
Several authorities consider the 1 mg/kg/d dose to be a LOAEL, but effects at the lowest dose were only observed in dams. Resulting US State RfDs range from 0.005 – 0.020 ug/kg-day (Post et al., 2021).

**Mouse**: Point of Departure = 4.35 µg/ml based on a serum PFOA benchmark concentration by New Jersey/New Hampshire (Post et al., 2021) for *lipid parameters/relative liver weight* in male mice from Loveless et al. (2006)

- Mouse to human toxicokinetic factor = 1 (Factor is not needed since BMD is based on serum concentration)
- Mouse to human toxicodynamic factor = 2.5 [IPCS (2005) default or 3 EPA (2014) default]
- Human toxicodynamic factor = 3 [default of IPCS (2005) and EPA (2014)].
- Human toxicokinetic factor = 8.4 [0.79 ml/day/kg arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) ÷ 0.094 ml/day/kg arithmetic 95% lower bound of sensitive group from Zhang et al. (2013, Table 2)]
- Database uncertainty factor = 1 (See footnote 7.)

RfD serum concentration = 0.058 µg/ml [4.35 µg/ml ÷ (1 x 3 x 3 x 8.4 x 1) = 0.058]

RfD = 0.01 µg/kg-day [0.058 µg/ml x 0.23 ml/day/kg [geometric mean clearance from Zhang et al. (2013, Table 2) assuming steady state]

Notes:
- It could be argued that a toxicodynamic UF of 0.1 could be applied for rodent to human differences in response to PPAR activation.

**Mouse**: Point of Departure = 0.3 mg/kg-day (10.4 µg/ml) NOAEL for *neonatal survival* found in Abbott et al. (2007)

- Mouse to human toxicokinetic factor = 1 (Factor is not needed since BMD is based on serum concentration)
- Mouse to human toxicodynamic factor = 2.5 [IPCS (2005) default or 3 EPA (2014) default]
- Human toxicodynamic factor = 3 [default of IPCS (2005) and EPA (2014)].
- Human toxicokinetic factor = 8.4 [0.79 ml/day/kg arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) ÷ 0.094 ml/day/kg arithmetic 95% lower bound of sensitive group from Zhang et al. (2013, Table 2)]
- Database uncertainty factor = 1 (See footnote 7)

RfD serum concentration = 0.14 µg/ml [10.4 µg/ml ÷ (1 x 3 x 3 x 8.4 x 1) = 0.14]

RfD = 0.03 µg/kg-day [0.14 µg/ml x 0.23 ml/day/kg [geometric mean clearance from
Mouse: Point of Departure = 0.94 mg/kg-day (no serum values available) NOAEL for immune suppression found in DeWitt et al. (2016).

Based on Lau et al. 2006, the serum level associated with in the mouse repeated dosing at 1 mg/kg-day is 23 µg/ml. Therefore, dosing at 0.94 mg/kg/d is estimated to be associated with a serum level of 22 µg/ml.

- Mouse to human toxicokinetic factor = 1 (Factor is not needed since BMD is based on serum concentration)
- Mouse to human toxicodynamic factor = 2.5 [IPCS (2005) default or 3 EPA (2014) default]
- Human toxicodynamic factor = 3 [default of IPCS (2005) and EPA (2014)].
- Human toxicokinetic factor = 8.4 [0.79 ml/day/kg arithmetic mean clearance of average group from Zhang et al. (2013, Table 2) ÷ 0.094 ml/day/kg arithmetic 95% lower bound clearance of sensitive group from Zhang et al. (2013, Table 2)]
- Database uncertainty factor = 1 (See footnote 7.)

- RfD serum concentration = 0.29 µg/ml [22 µg/ml ÷ (1 x 3 x 3 x 8.4 x 1) = 0.29]
- RfD = 0.07 µg/kg-day [0.29 µg/ml x 0.23 ml/day/kg [geometric mean clearance from Zhang et al. (2013, Table 2) assuming steady state]